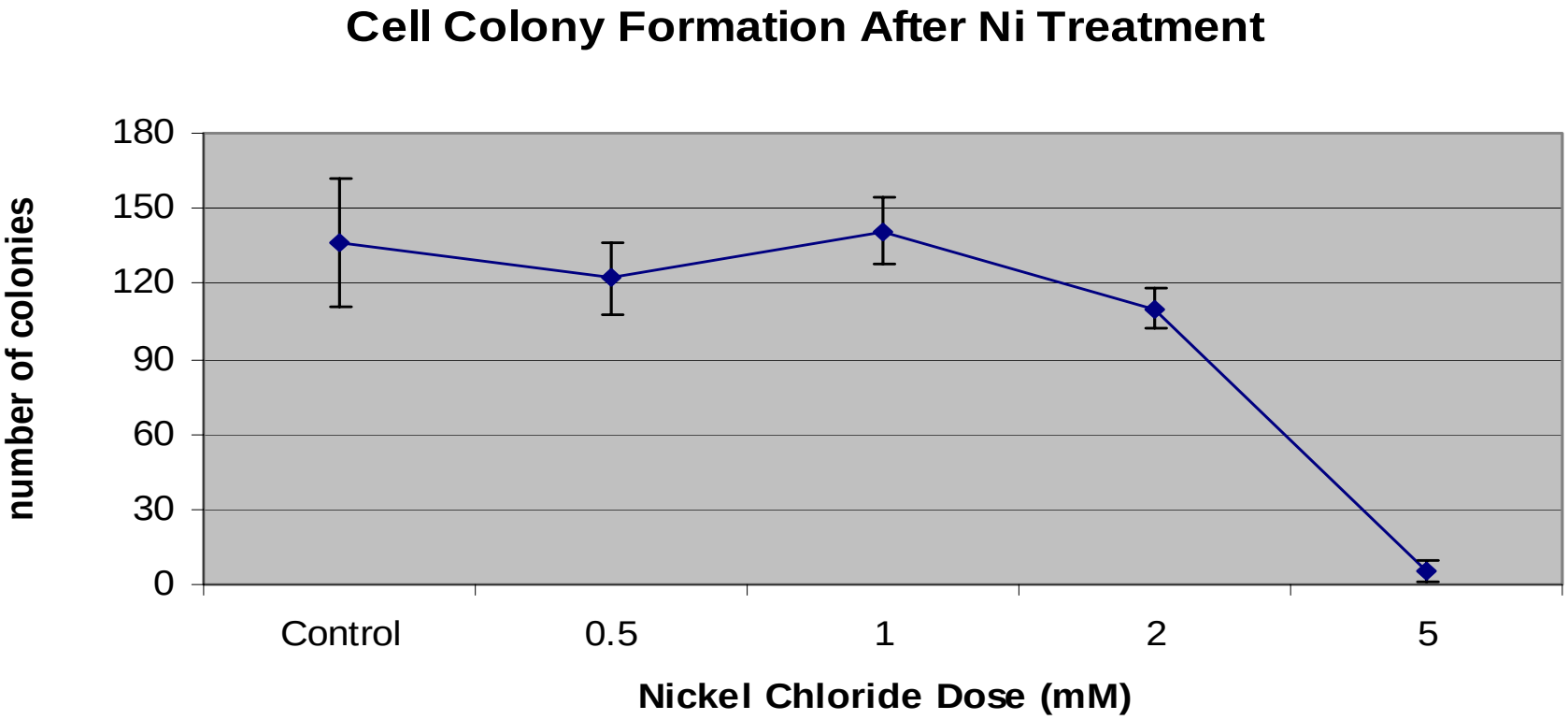
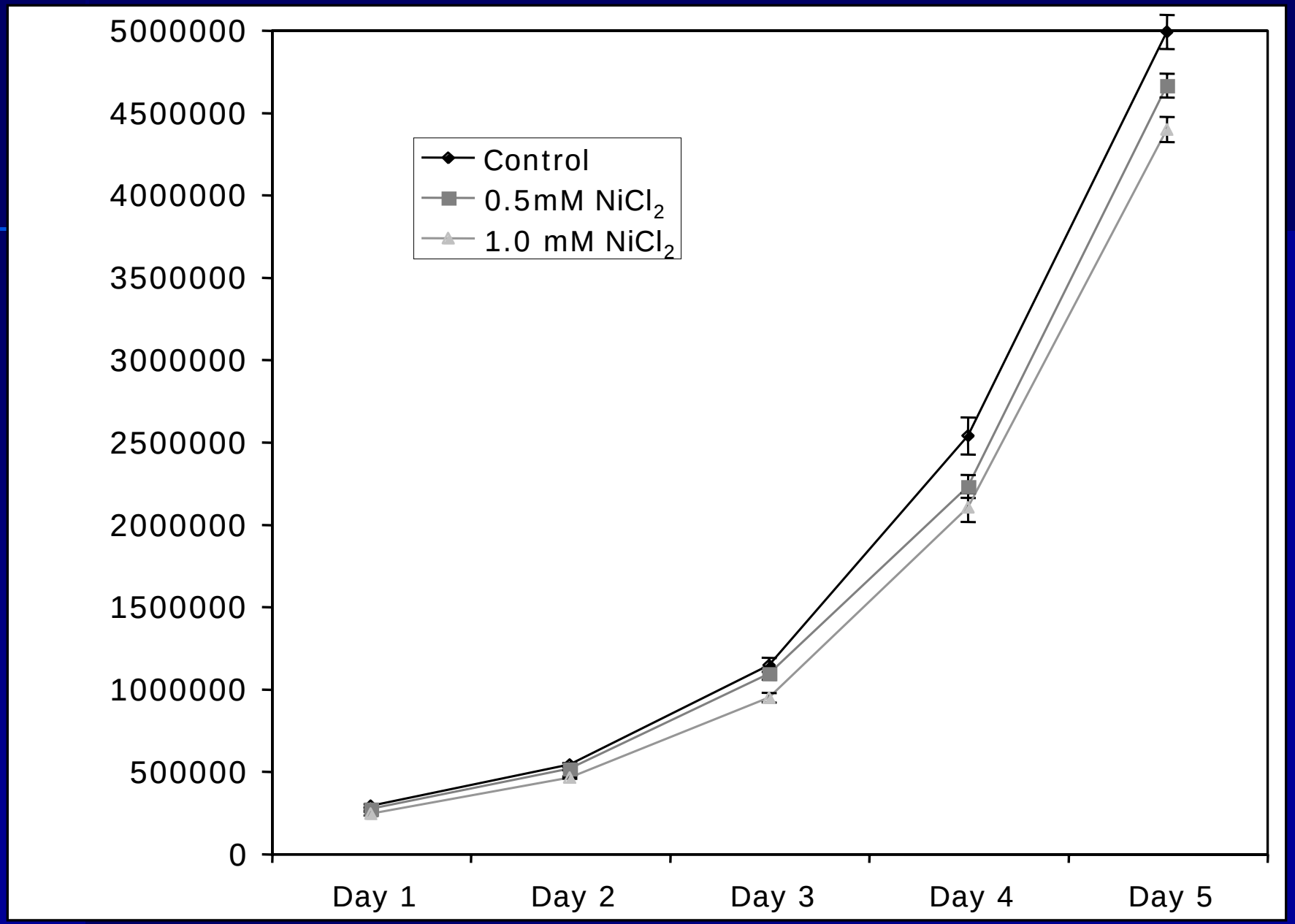
Nickel and Megasites

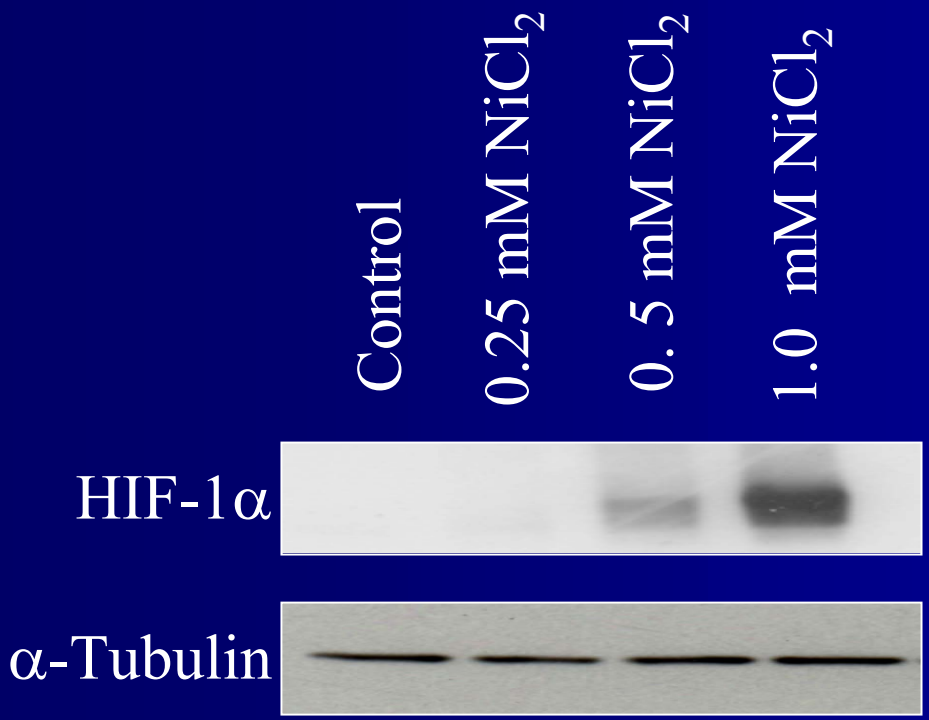
- Nickel compounds are major contaminants in many superfund sites
- Although Ni ions are required for certain enzymes in bacteria and plants (Ureases, Dehydrogenases), No known function in mammals.
- Certain Ni compounds that deliver Ni ions into cells, are potently carcinogenic (nasal, lung cancers etcat site of exposure)
- Additional toxicities include: contact dermatitis, depression of lung function, and cardiovascular effects.

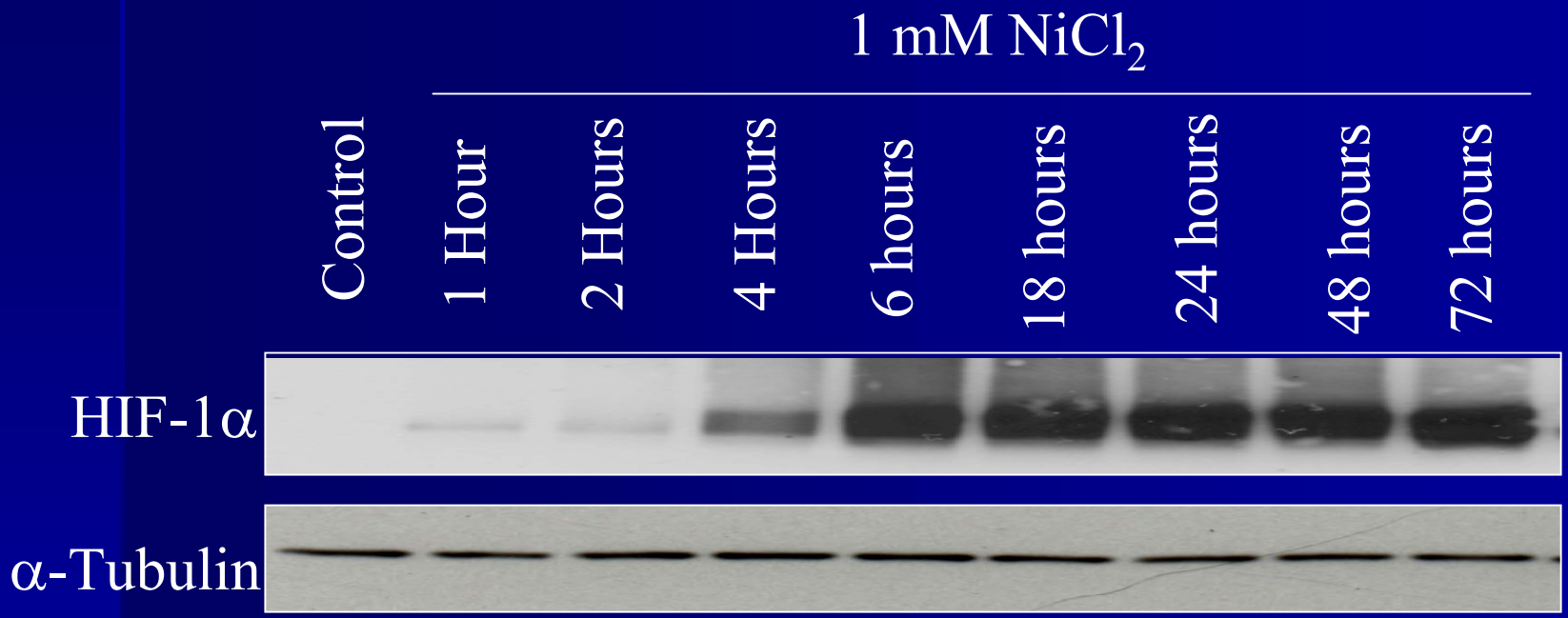
Effect of 24 hr exposure of A549 Cells to $NiCl_2$ On Cell Colony Formation




5000000
4500000
4000000
3500000
3000000
2500000
2000000
1500000
1000000
500000
0
Control
0.5mM NiCl2
1.0 mM NiCl2
Day 1
Day 2
Day 3
Day 4
Day 5

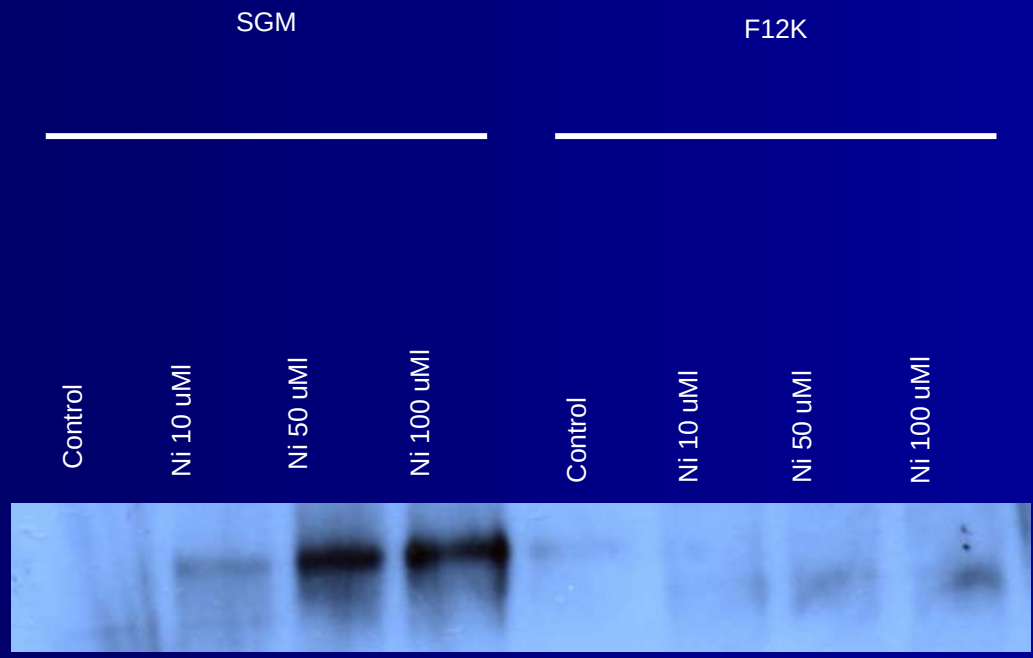
HIF-1 alpha protein levels at 5 hrs

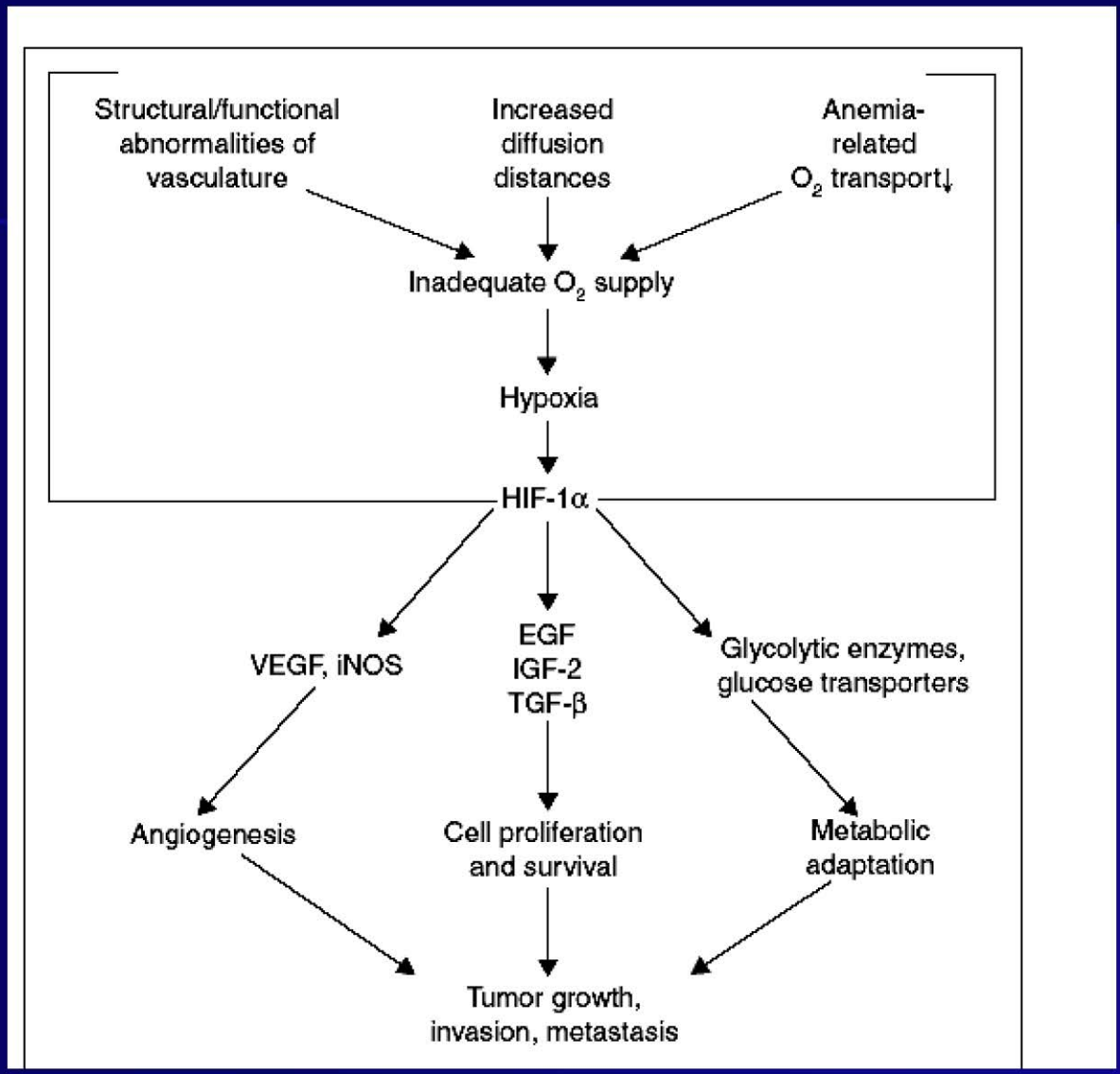



1 mM $NiCl_2$
Control
1 Hour
2 Hours
4 Hours
6 hours
18 hours
24 hours
48 hours
72 hours
HIF-1α
α-Tubulin

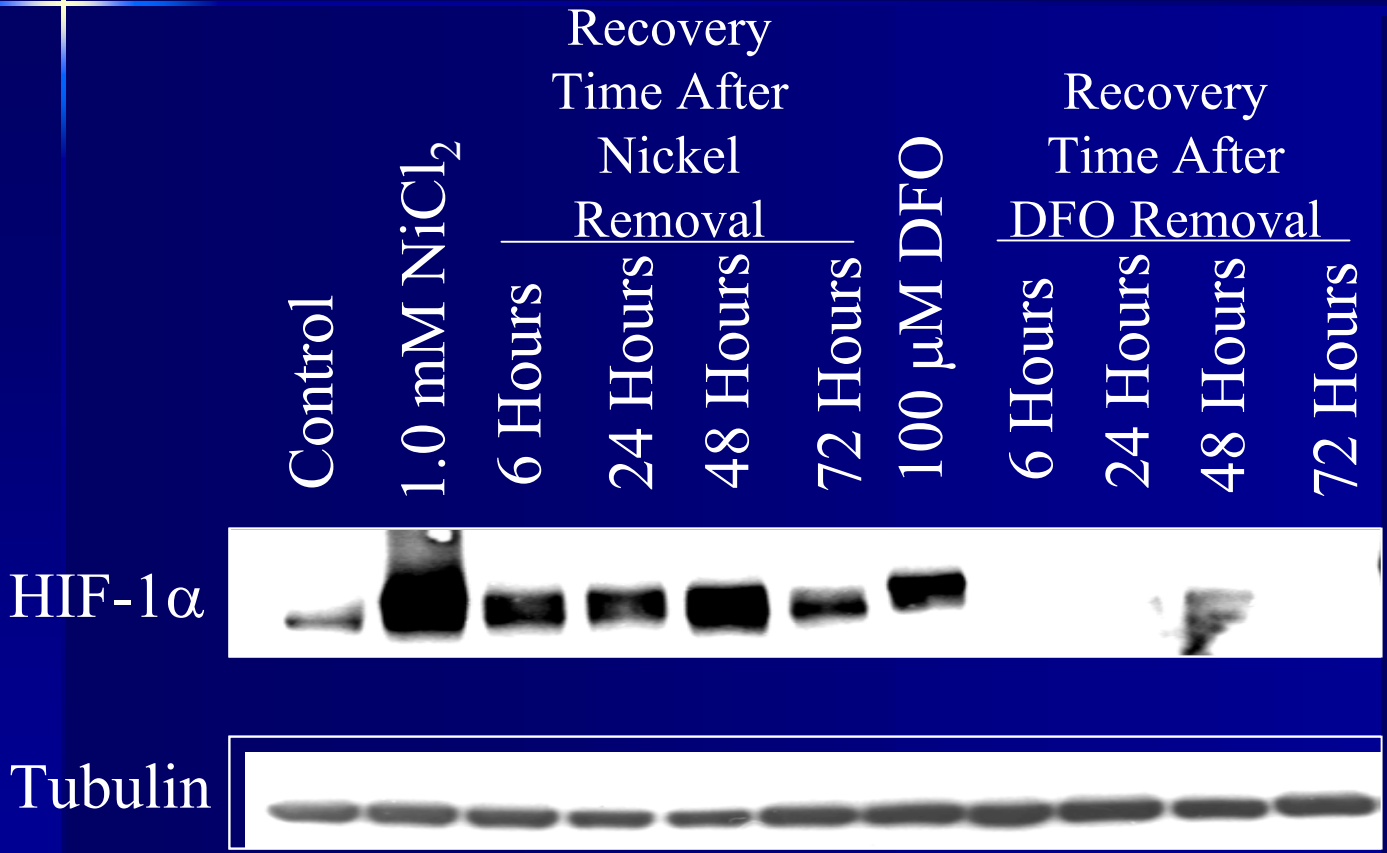
A549

5 hr exp.

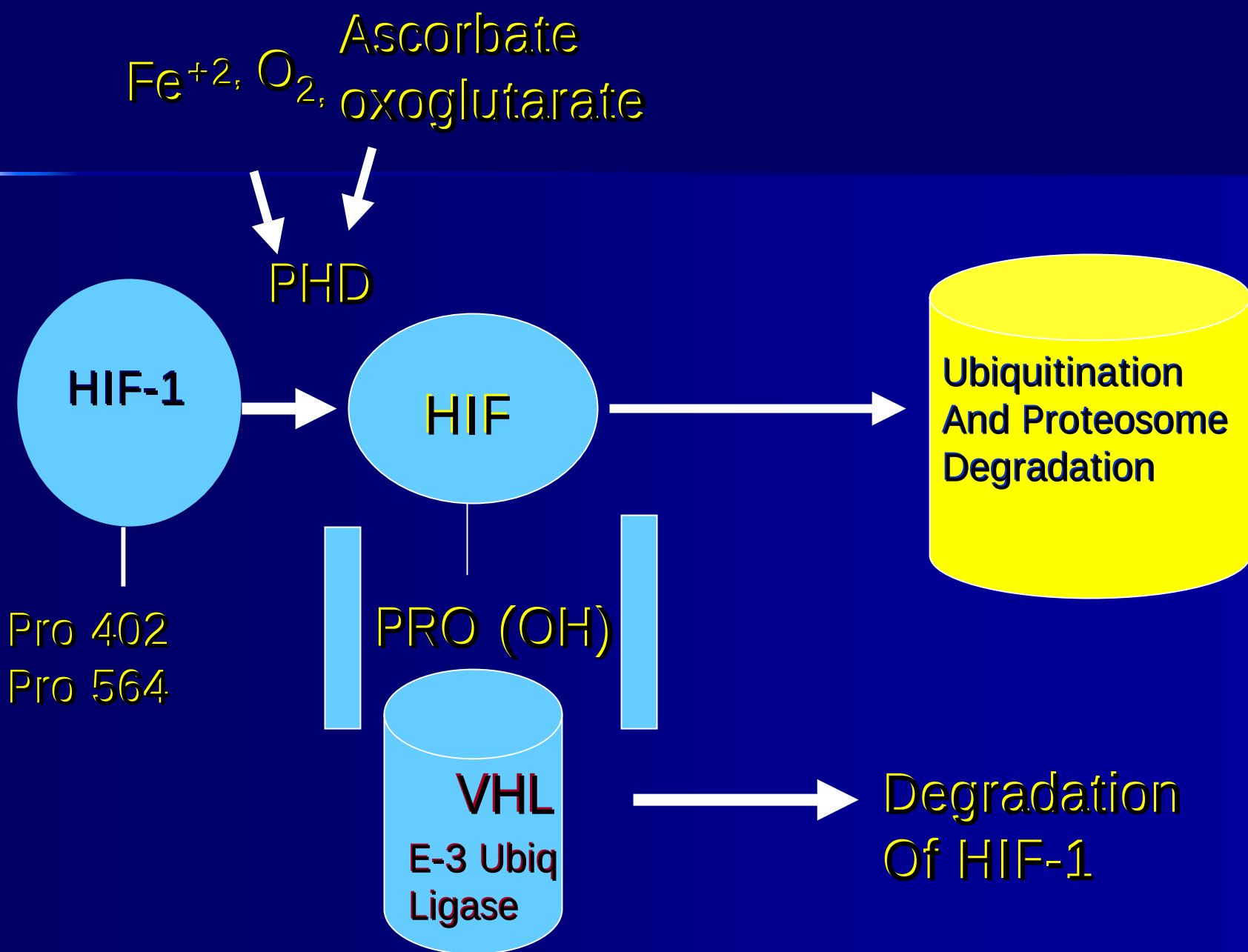

SGM
F12K
Control
Ni 10 uMl
Ni 50 uMl
Ni 100 uMl
Control
Ni 10 uMl
Ni 50 uMl
Ni 100 uMl


Structural/functional abnormalities of vasculature
Increased diffusion distances
Anemia-related O_2 transport↓
Inadequate O_2 supply
Hypoxia
HIF-1α
VEGF, iNOS
EGF
IGF-2
TGF-β
Glycolytic enzymes, glucose transporters
Angiogenesis
Cell proliferation and survival
Metabolic adaptation
Tumor growth, invasion, metastasis

Persistent Stabilization of HIF --1 alpha by $NiCl_2$

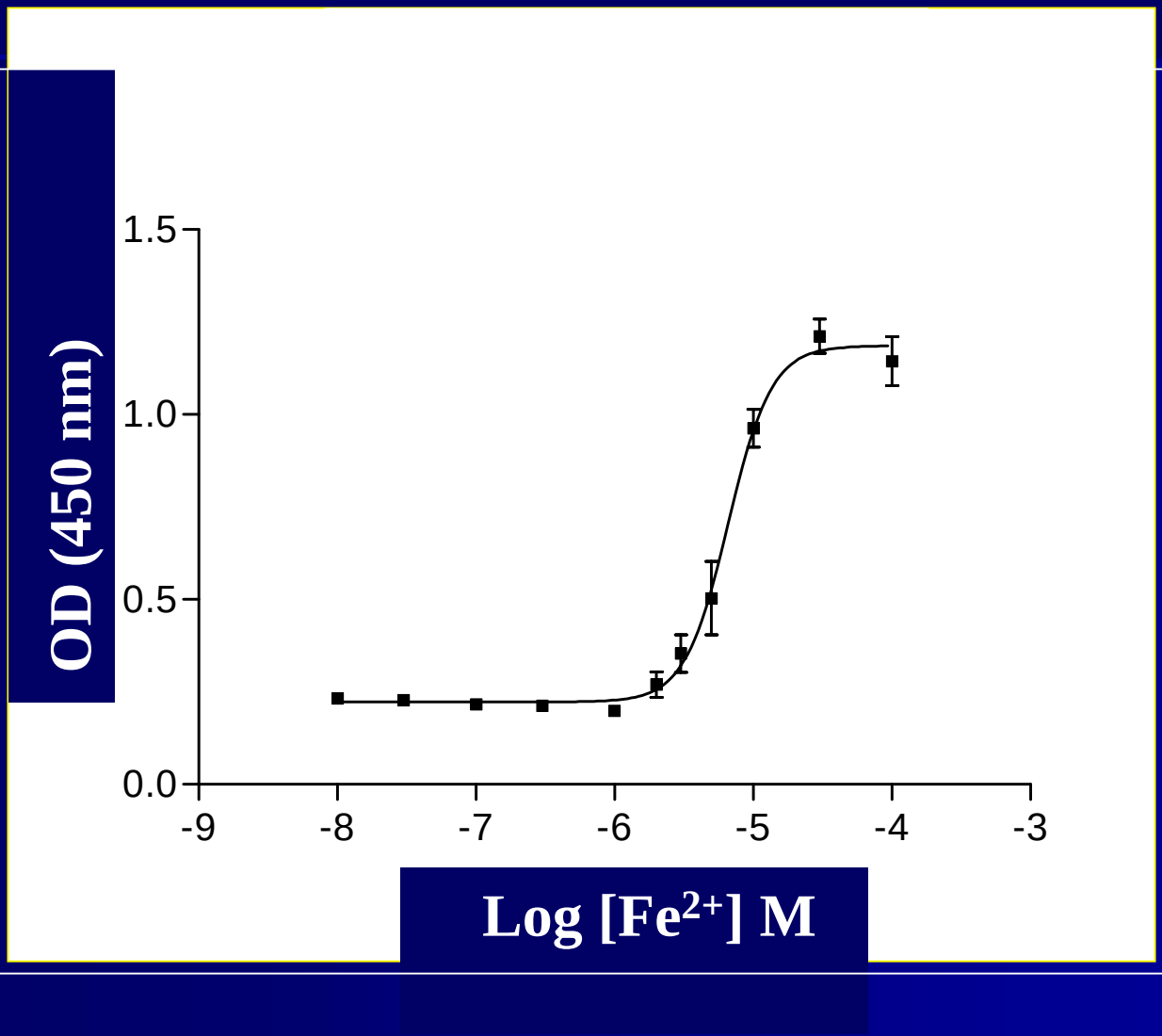


Prolyl Hydroxylase Regulates Hif-1 stability

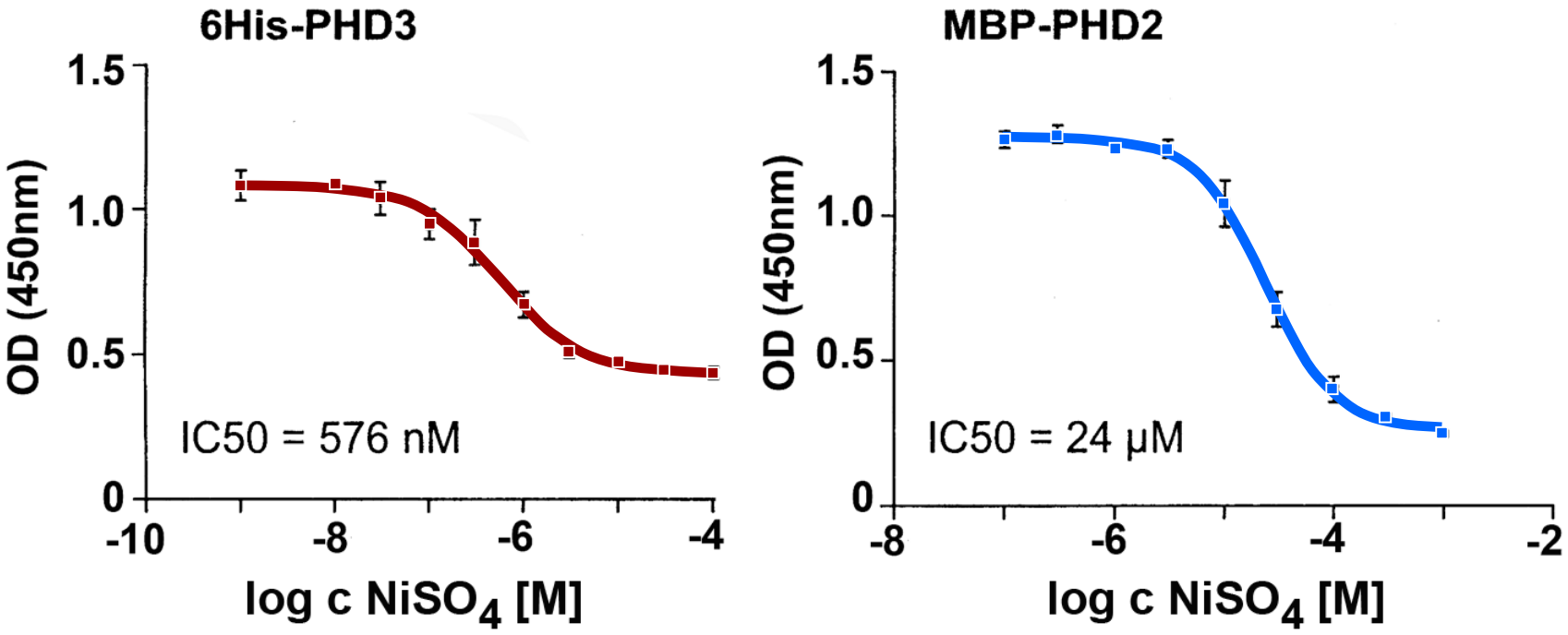


Lack of reversibility of Ni Inhibition of HIF hydroxylation

Requirement for Fe of PH2 (2ug protein)

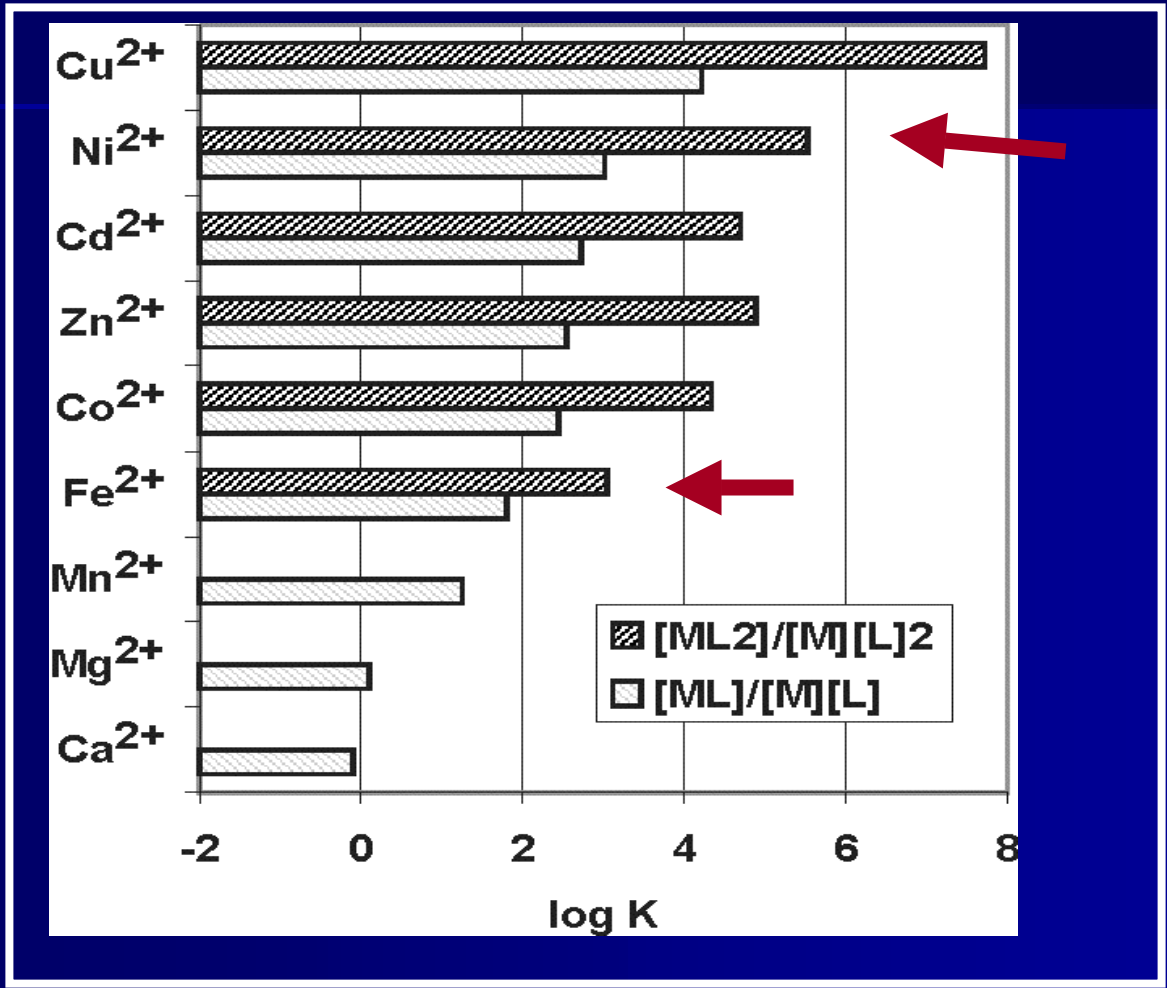


Inhibition of HIF-PHD2 and HIF-PHD3 by $NiSO_4$

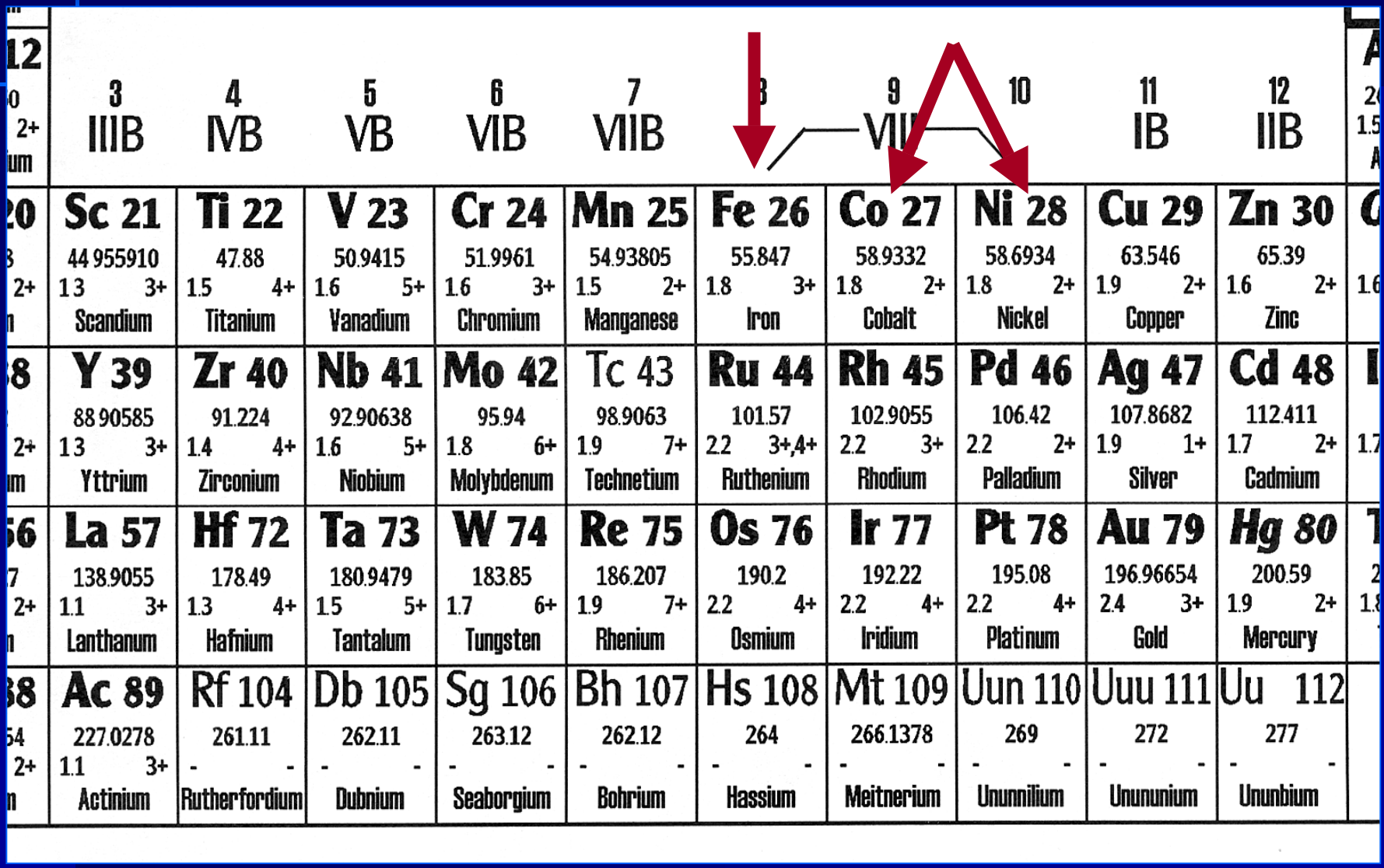


Biochemical assay performed according to Oehme et al. (2004) Anal. Biochem. **330**, 74 - 80
6His-PHD3: expressed in insect cells, initially purified by incubation with DEAE-Sepharose
MBP-PHD2: expressed in E. coli, purified by affinity chromatography with Amylose Resin
MBP-PHD2: 2 µg 6His-PHD3: 20 ng (estimated)
Measurements were performed in triplicate and are show as mean values ± SEM

Binding Constants of Metal Ions to Immidazole



Periodic Table

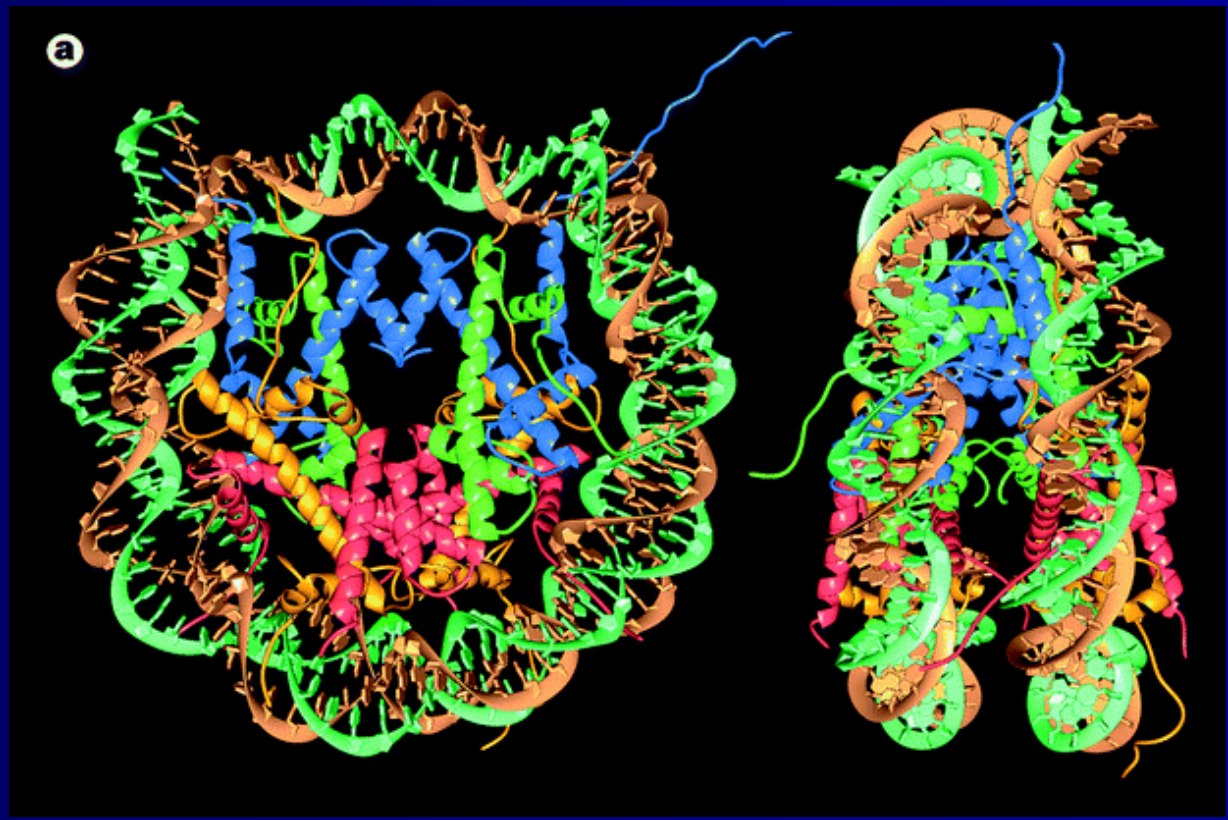


3 IIIB	4 IVB	5 VB	6 VIB	7 VIIB	8	9 VIII	10	11 IB	12 IIB
Sc 21	Ti 22	V 23	Cr 24	Mn 25	Fe 26	Co 27	Ni 28	Cu 29	Zn 30
44.955910	47.88	50.9415	51.9961	54.93805	55.847	58.9332	58.6934	63.546	65.39
1.3 3+	1.5 4+	1.6 5+	1.6 3+	1.5 2+	1.8 3+	1.8 2+	1.8 2+	1.9 2+	1.6 2+
Scandium	Titanium	Vanadium	Chromium	Manganese	Iron	Cobalt	Nickel	Copper	Zinc
Y 39	Zr 40	Nb 41	Mo 42	Tc 43	Ru 44	Rh 45	Pd 46	Ag 47	Cd 48
88.90585	91.224	92.90638	95.94	98.9063	101.57	102.9055	106.42	107.8682	112.411
1.3 3+	1.4 4+	1.6 5+	1.8 6+	1.9 7+	2.2 3+,4+	2.2 3+	2.2 2+	1.9 1+	1.7 2+
Yttrium	Zirconium	Niobium	Molybdenum	Technetium	Ruthenium	Rhodium	Palladium	Silver	Cadmium
La 57	Hf 72	Ta 73	W 74	Re 75	Os 76	Ir 77	Pt 78	Au 79	Hg 80
138.9055	178.49	180.9479	183.85	186.207	190.2	192.22	195.08	196.96654	200.59
1.1 3+	1.3 4+	1.5 5+	1.7 6+	1.9 7+	2.2 4+	2.2 4+	2.2 4+	2.4 3+	1.9 2+
Lanthanum	Hafnium	Tantalum	Tungsten	Rhenium	Osmium	Iridium	Platinum	Gold	Mercury
Ac 89	Rf 104	Db 105	Sg 106	Bh 107	Hs 108	Mt 109	Uun 110	Uuu 111	Uu 112
227.0278	261.11	262.11	263.12	262.12	264	266.1378	269	272	277
1.1 3+	- -	- -	- -	- -	- -	- -	- -	- -	- -
Actinium	Rutherfordium	Dubnium	Seaborgium	Bohrium	Hassium	Meitnerium	Ununnilium	Unununium	Ununbium

Summary of Findings

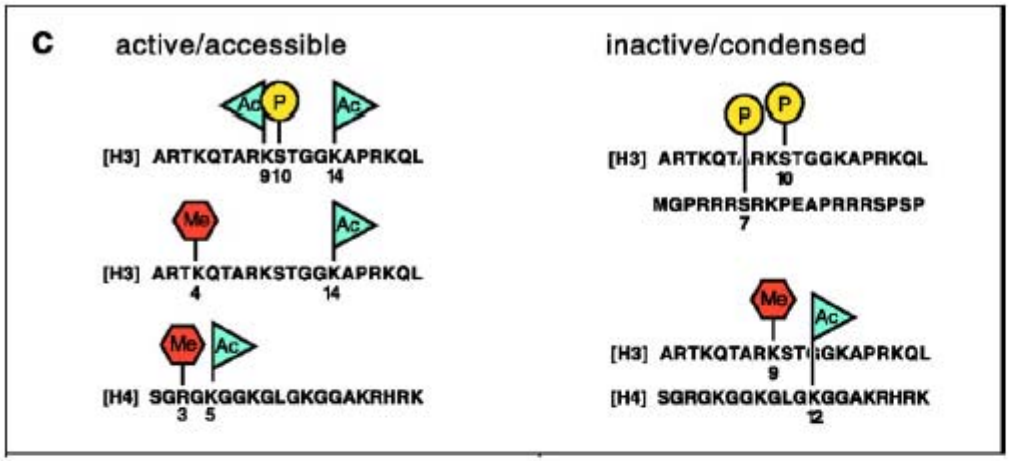
- Transformation of Chinese hamster primary embryo fibroblast by Nickel Compounds inactivated tumor suppressor gene by DNA Methylation (*Science* 251:796--799 (1991)
- Carcinogenic Nickel compounds Induce trangene silencing based upon the location of the trangene near Heterochromatin (mammalian cells) or a telomere silencing in yeast. (Lee et al MCB 15 2547 1995)

Structure of Nucleosome



Luger K et al. Nature, **389:**251

Blue: H3; green: H4; yellow: H2A; red: H2B



Jenuwein and Allis, Science, August 2001.

Histone Code Hypothesis:
Different combinations of histone modifications, especially located near or within a gene's promoter, may be VERY SPECIFIC to the transcriptional state of that gene.

Associated with active/accessible chromatin

H3K9 Acetylated
H3K14 Acetylated
H3K4 (di-)Methylated
H4 Acetylated

Associated with inactive/condensed chromatin

H3K9 Methylated
H3K9 di-Methylated (inactive X-chromosome)
H3K27 tri-Methylated (inactive X-chromosome)
H3K9 tri-Methylated (pericentromeric heterochromatin)
H3K27 mono-Methylated (pericentromeric heterochromatin)

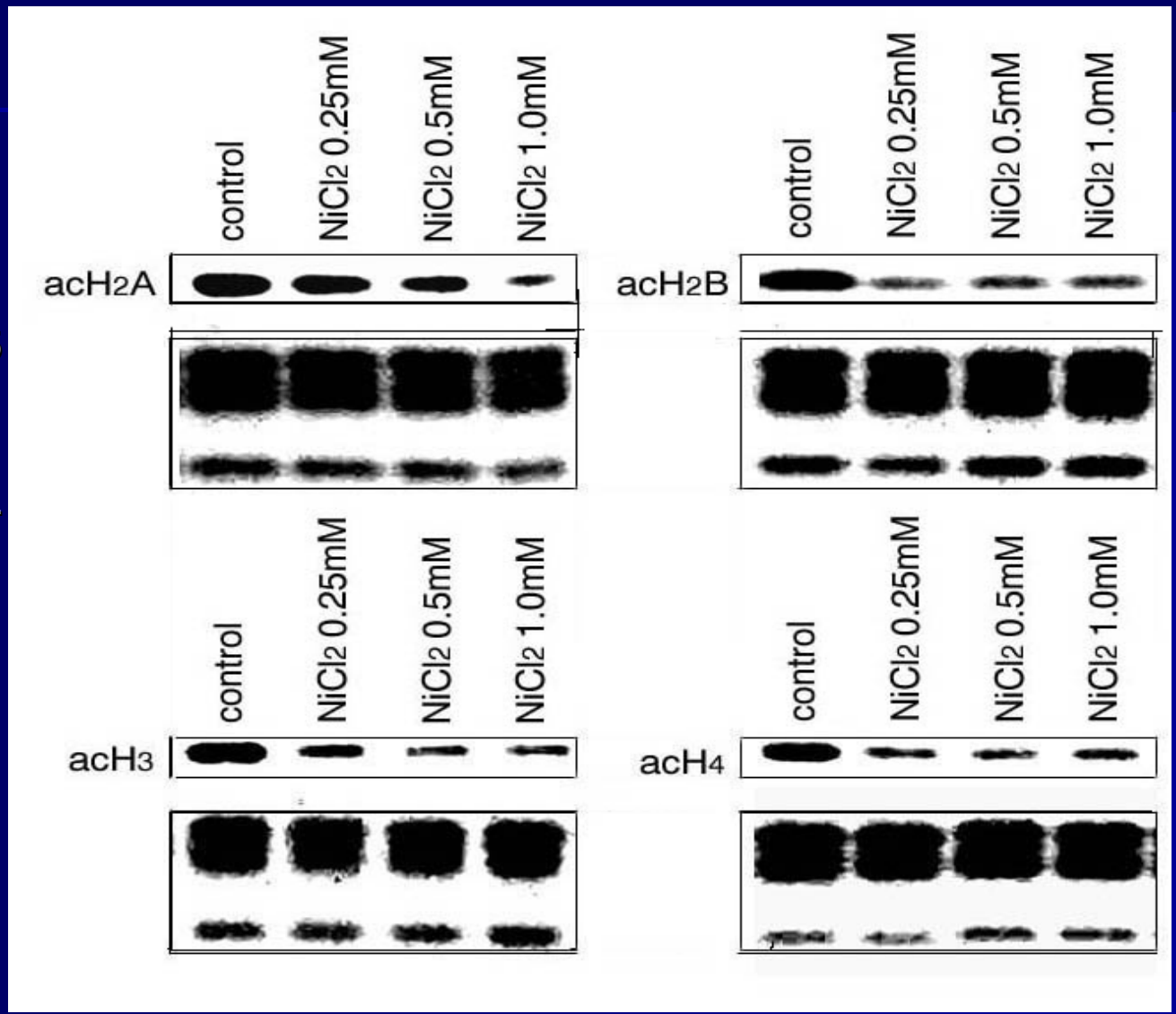
ADD IN DNA METHYLATION, AND THE TRANSCRIPTIONAL REGULATION OF A GENE CAN BECOME VERY COMPLEX!!!

Aim1

Fig.1A. Ni decreases histone acetylation in A549 cells.

A549
24h

H3 15
H2A 14
H2B 13
H4 11



Aim1

Fig.1B. Ni decreases histone acetylation in other cell lines.

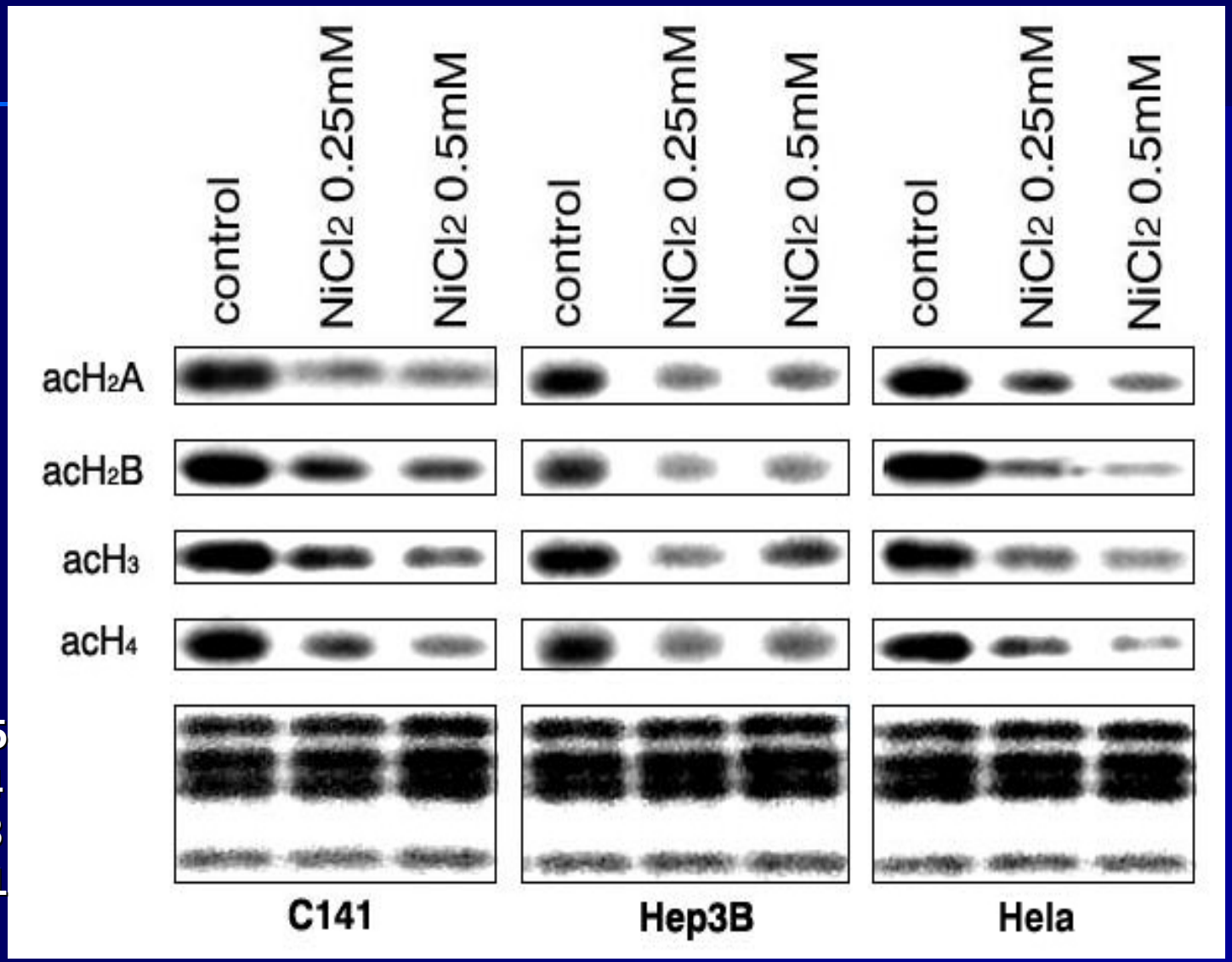


Table 1
Exposure to TSA reverted the ability of Ni-transformed cells to grow in soft agar

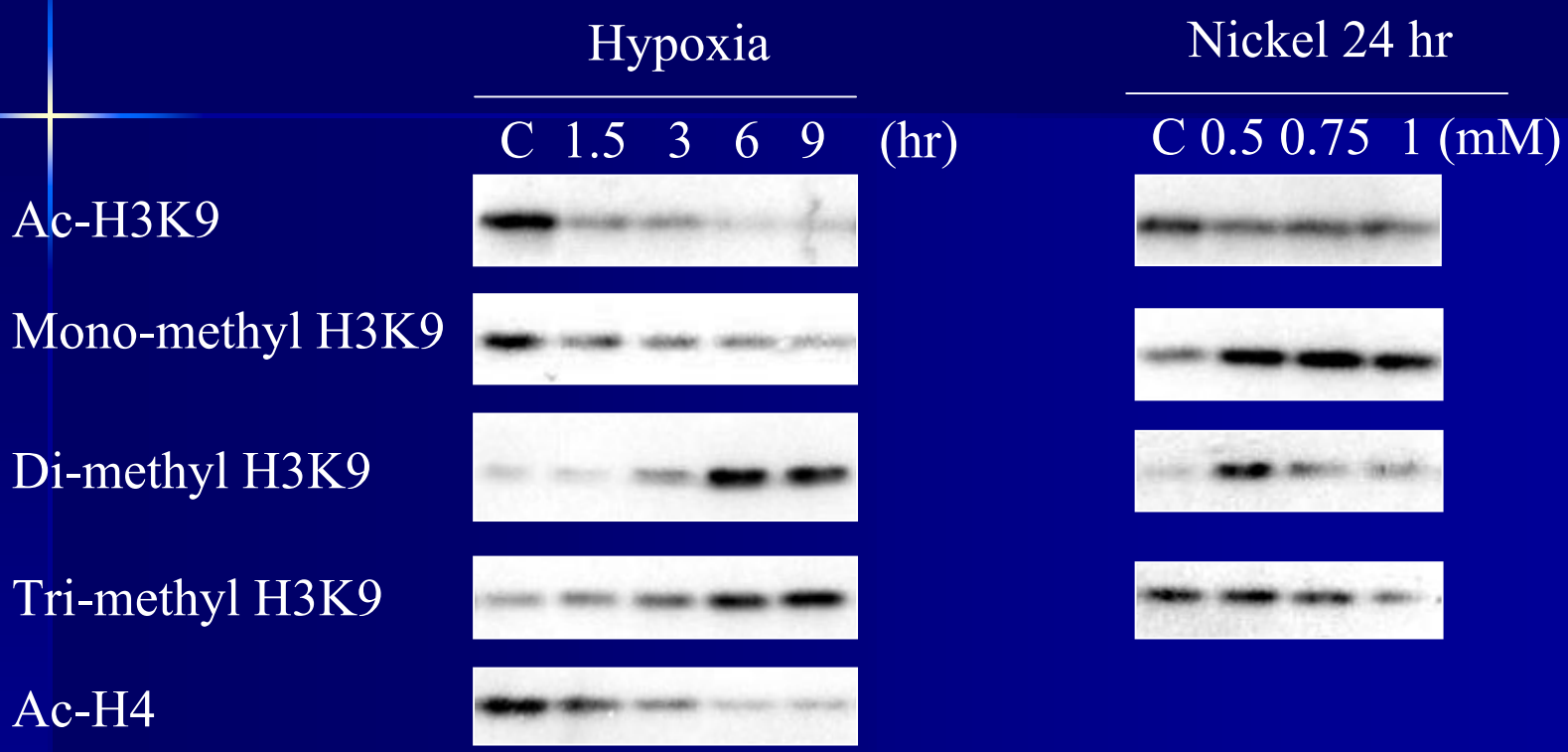
Ni-transformed clones	0 ng/ml TSA	5 ng/ml TSA	25 ng/ml TSA
1	100.0 ± 2.7	38.5 ± 5.2**	16.0 ± 1.3**
2	100.0 ± 12.4	65.7 ± 11.0*	34.2 ± 1.4**
3	100.0 ± 6.4	54.1 ± 6.9**	22.3 ± 0.3**
4	100.0 ± 13.0	48.7 ± 5.5**	36.5 ± 5.9**
5	100.0 ± 0.85	33.3 ± 1.1**	21.6 ± 4.1**

Note. One million of the Ni-transformed cells were seeded into flasks and exposed to 0, 5, or 25 ng/ml TSA for 24 h. The TSA containing medium was removed and cells were rinsed three times with saline A. Fresh medium was added and the cells were allowed to grow to ~80% confluence. The cultures were split and the cells were seeded at a density of 1 million cells again. And cells were treated with TSA from a second time. After third round of TSA exposure, cells were allowed to repopulate the culture prior to test for the ability of anchorage-independent growth in soft agar. Results are expressed as a percentage of control (0 ng/ml TSA treatment group). Values are mean ± SE. Asterisk indicates significant differences from that of control.

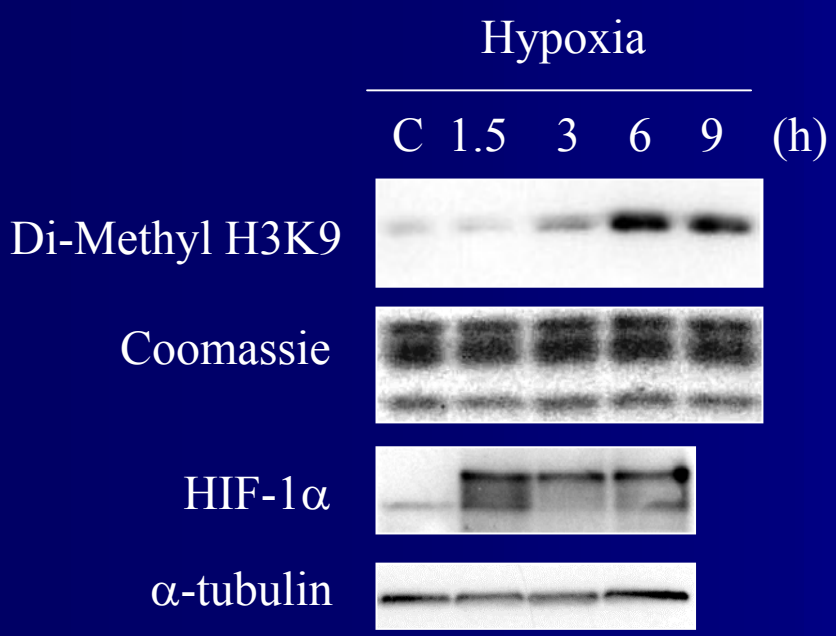
* $P < 0.05$.

** $P < 0.005$, $N = 3$.

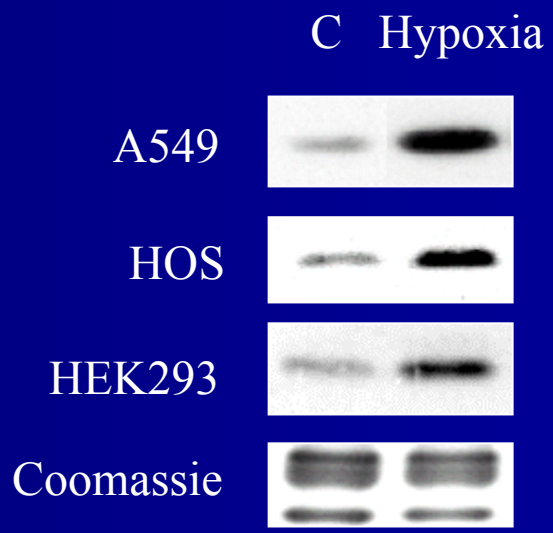
A549


Hypoxia
C 1.5 3 6 9 (hr)
Nickel 24 hr
C 0.5 0.75 1 (mM)
Ac-H3K9
Mono-methyl H3K9
Di-methyl H3K9
Tri-methyl H3K9
Ac-H4

a


Hypoxia
C 1.5 3 6 9 (h)
Di-Methyl H3K9
Coomassie
HIF-1α
α-tubulin

b


C Hypoxia
A549
HOS
HEK293
Coomassie

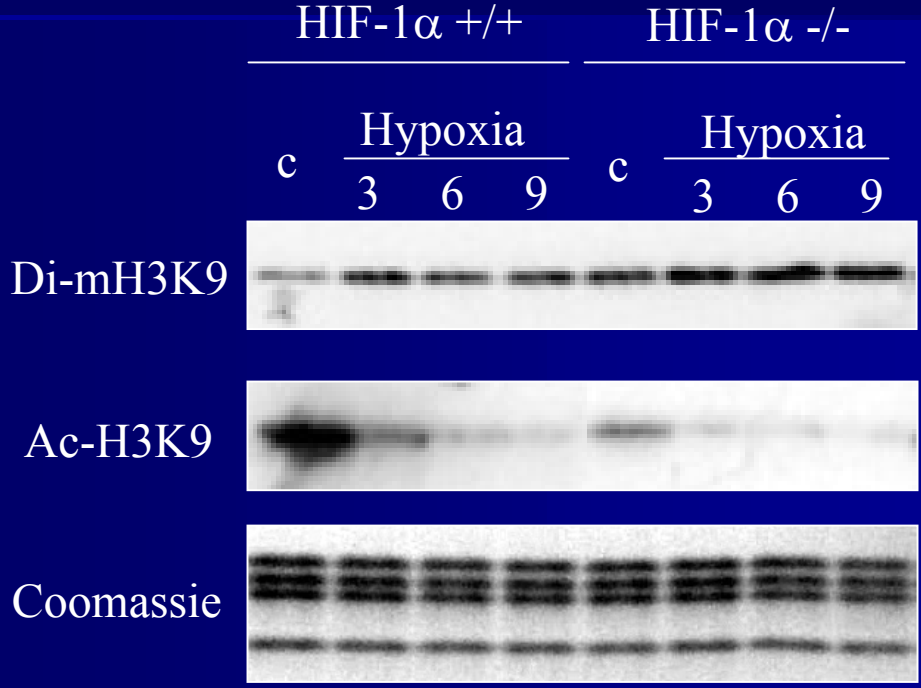
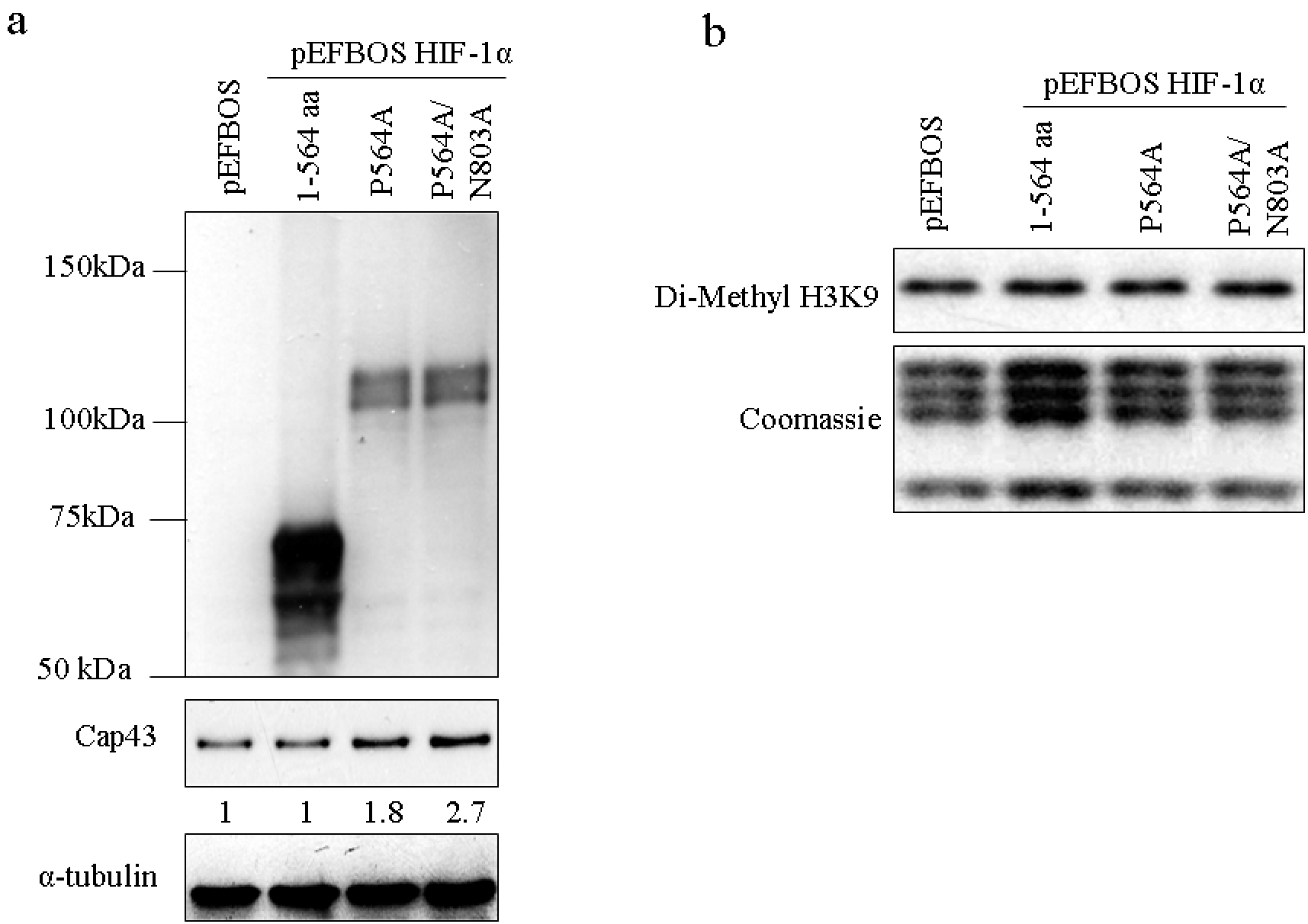
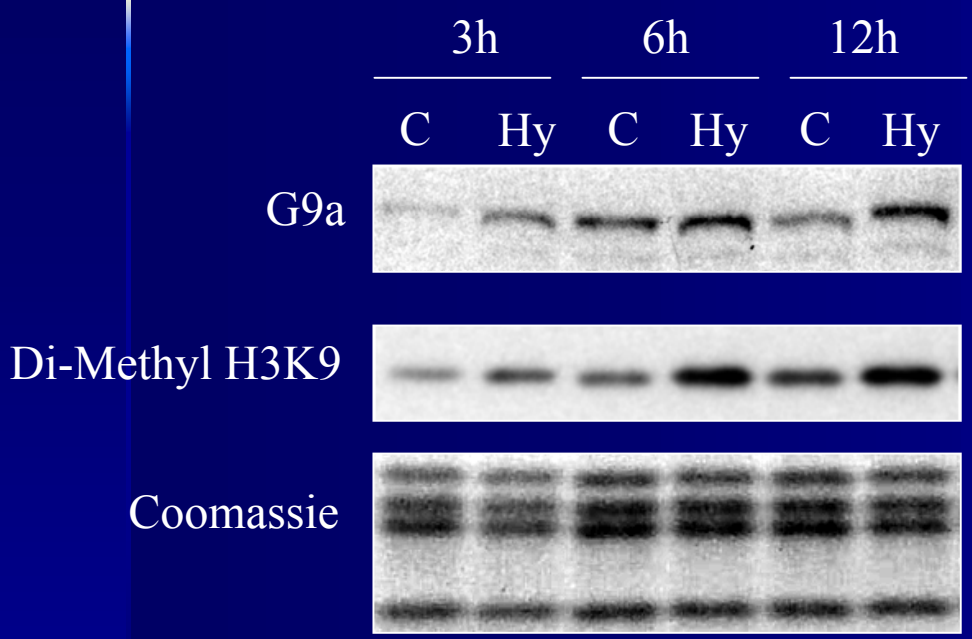

HIF-1α +/+
HIF-1α -/-
c
Hypoxia
3
6
9
c
Hypoxia
3
6
9
Di-mH3K9
Ac-H3K9
Coomassie

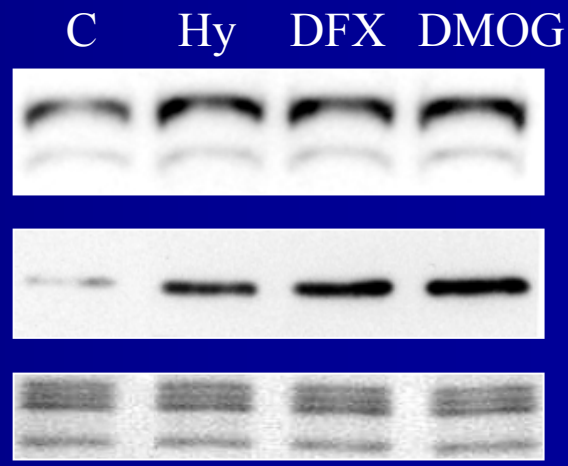
Figure 4


a
pEFBOS
pEFBOS HIF-1α
1-564 aa
P564A
P564A/
N803A
150kDa
100kDa
75kDa
50 kDa
Cap43
1
1
1.8
2.7
α-tubulin
b
pEFBOS
pEFBOS HIF-1α
1-564 aa
P564A
P564A/
N803A
Di-Methyl H3K9
Coomassie

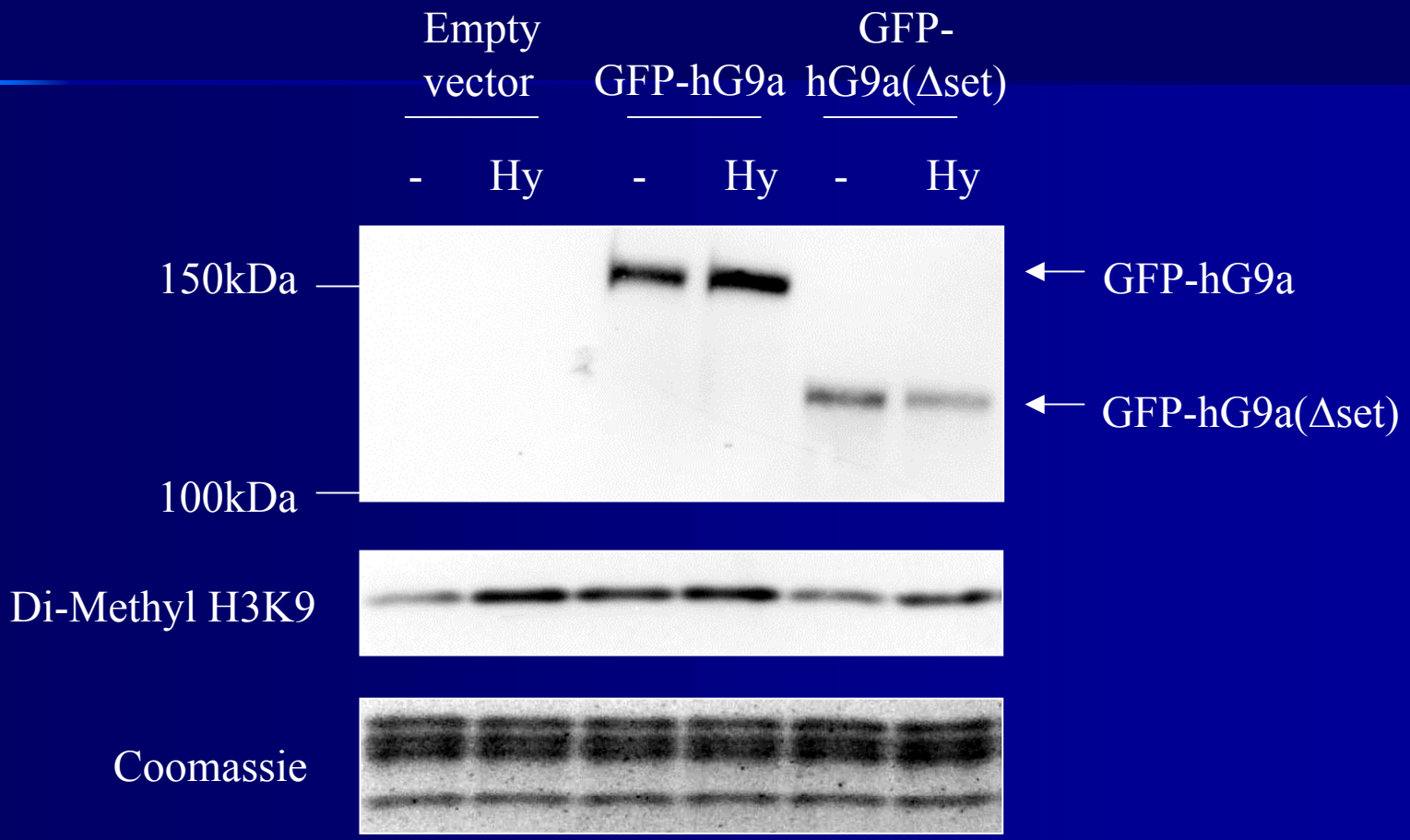
a


3h
6h
12h
C
Hy
C
Hy
C
Hy
G9a
Di-Methyl H3K9
Coomassie

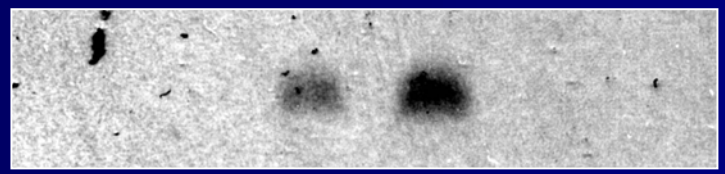
b


C
Hy
DFX
DMOG

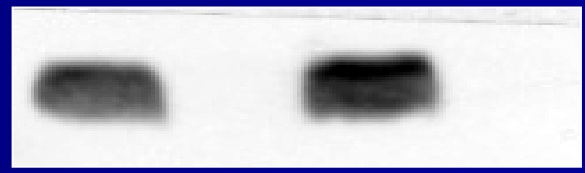
a


Empty vector
GFP-hG9a
GFP-hG9a(Δset)
- Hy - Hy - Hy
150kDa
100kDa
GFP-hG9a
GFP-hG9a(Δset)
Di-Methyl H3K9
Coomassie

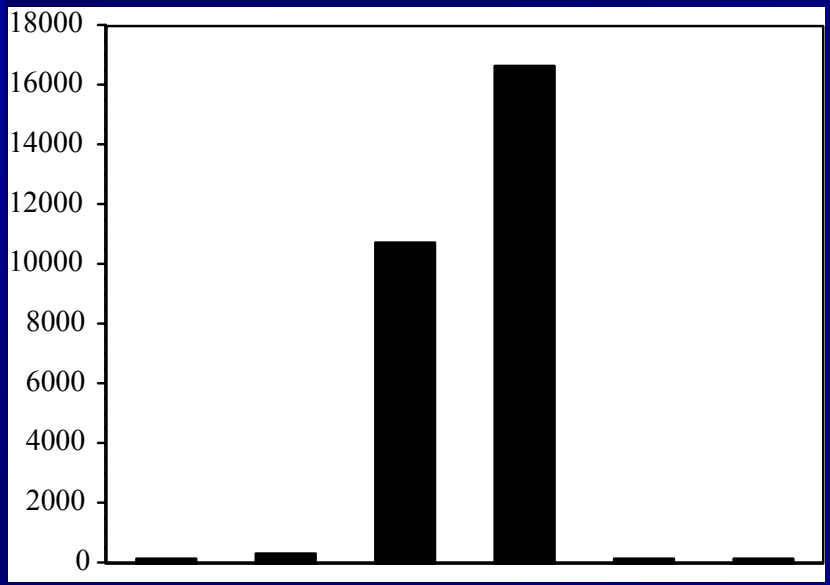
b

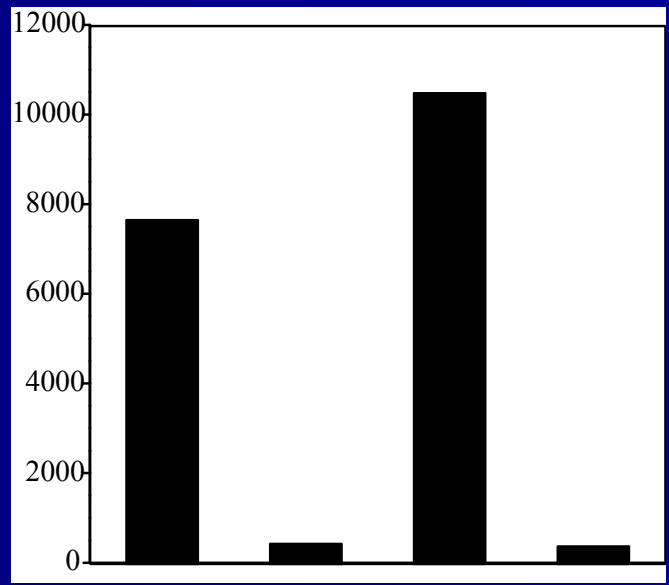


Flurogram

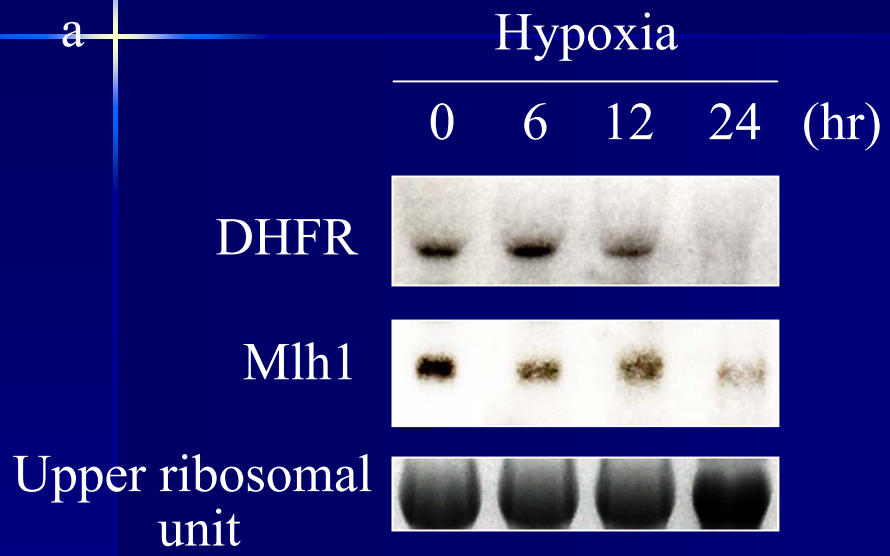


CPM

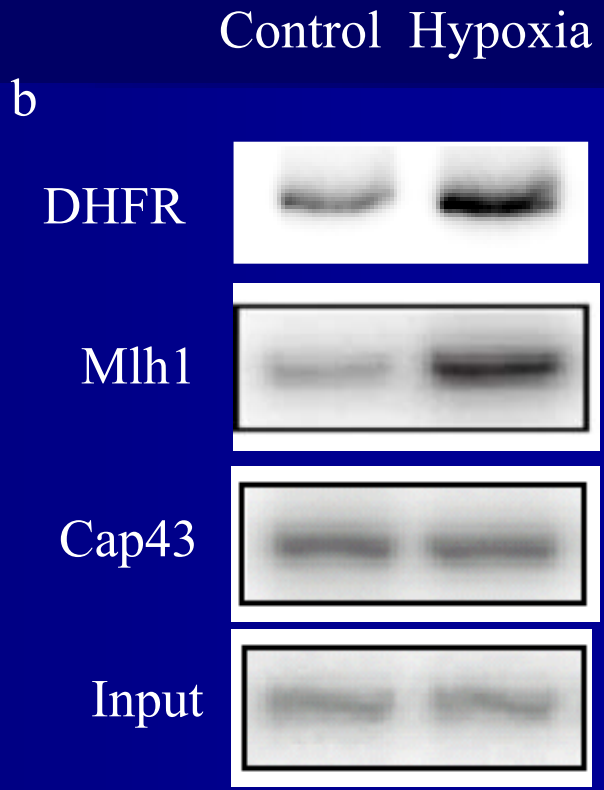




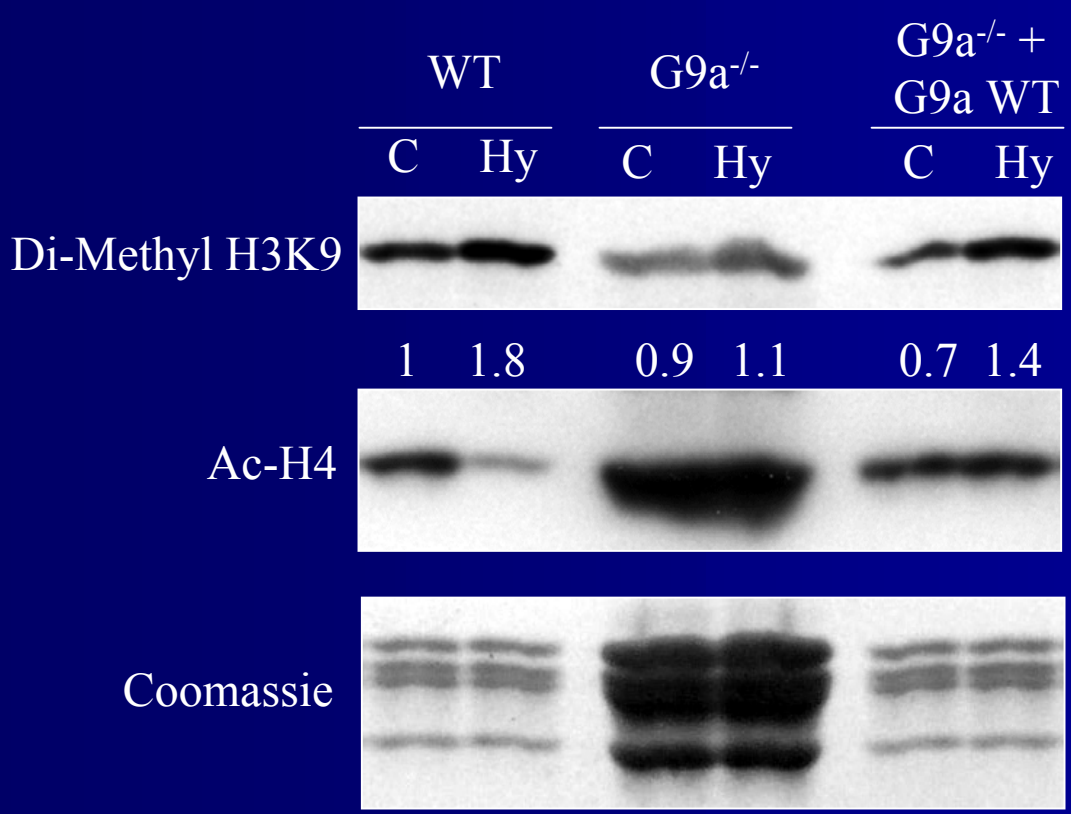
a


Hypoxia
0
6
12
24
(hr)
DHFR
Mlh1
Upper ribosomal unit

b

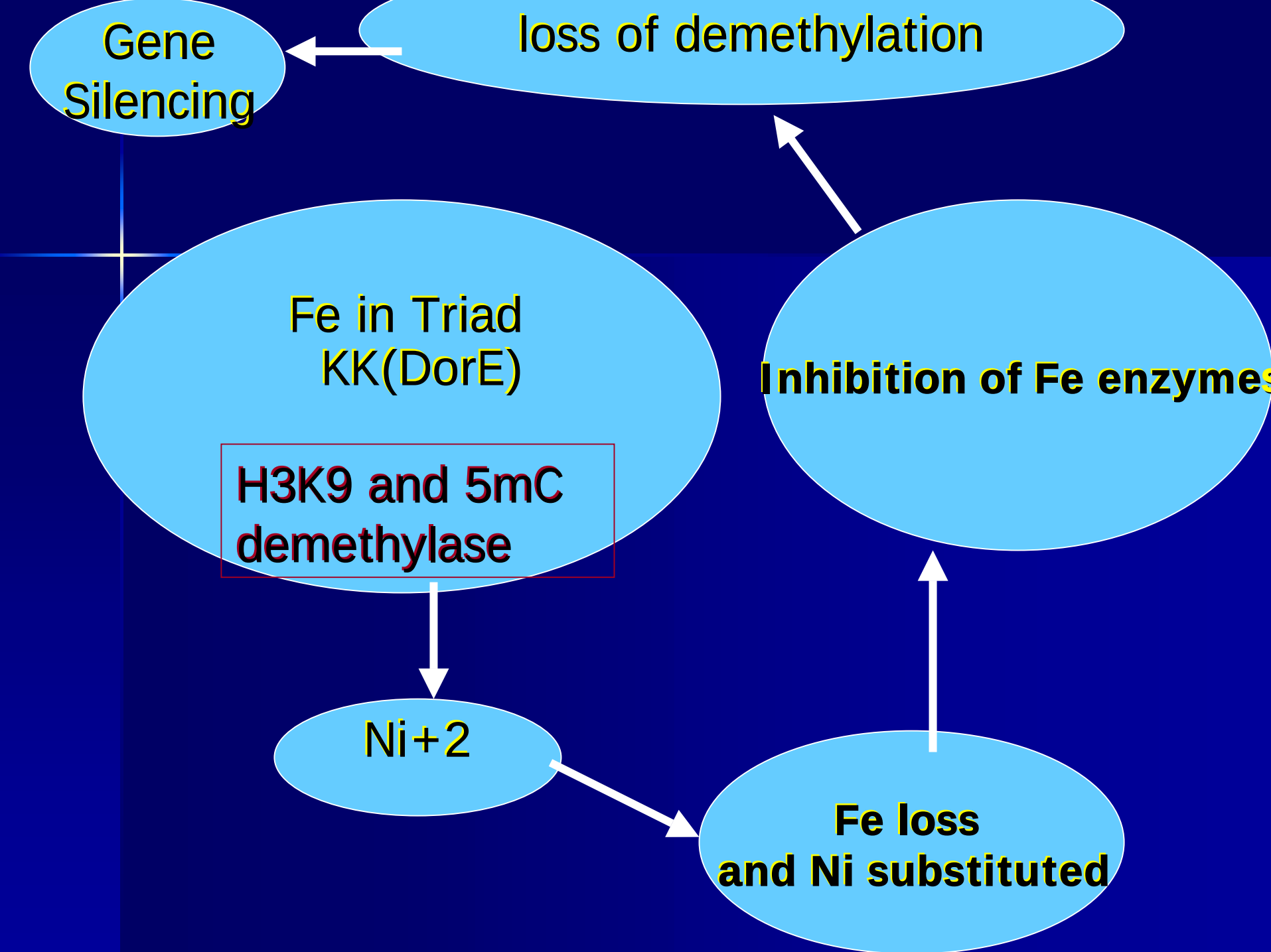

Control
Hypoxia
DHFR
Mlh1
Cap43
Input

a


WT
G9a-/-
G9a-/- + G9a WT
C
Hy
C
Hy
C
Hy
Di-Methyl H3K9
1
1.8
0.9
1.1
0.7
1.4
Ac-H4
Coomassie

Summary of the Effect of Ni compounds on G9a and H3K9 methylation

- Ni ions inhibited G9a activity and decreased its presence in the nucleus (see poster by H. Chen)
- We have been working on Fe, Oxoglutarate, ascorbic acid dependent H3K9 demethylase and 5-methylctyosine- demethylase and have found theseactivities in crude cell extracts. Ni ions are effective inhibitors of these enzymes.
- These are new enzymes and further work on their identification, purification and characterization is required.


Gene Silencing
loss of demethylation
Fe in Triad KK(DorE)
H3K9 and 5mC demethylase
Inhibition of Fe enzymes
Ni+2
Fe loss and Ni substituted

Ackowledgements

Todd Davison
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Karin Ke
Thomas Kluz
Konstatin Salnikow
Qin Li
Yan Yan

Randy Johnson
Yoichi Shinkai